This article was downloaded by:

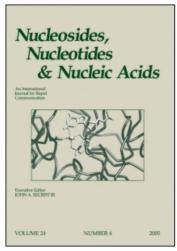
On: 27 January 2011

Access details: Access Details: Free Access

Publisher Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-

41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597286

Synthesis of 4'-(Hydroxymethyl)Guanosine and a Phosphonate Analogue of Guanylic Acid

John C. Martin^a; Julien P. H. Verheyden^a Syntex Research, Palo Alto, CA

To cite this Article Martin, John C. and Verheyden, Julien P. H.(1988) 'Synthesis of 4'-(Hydroxymethyl)Guanosine and a Phosphonate Analogue of Guanylic Acid', Nucleosides, Nucleotides and Nucleic Acids, 7: 3, 365 — 374

To link to this Article: DOI: 10.1080/07328318808068716 URL: http://dx.doi.org/10.1080/07328318808068716

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

SYNTHESIS OF 4'-(HYDROXYMETHYL)GUANOSINE AND A PHOSPHONATE ANALOGUE OF GUANYLIC ACID 1

John C. Martin² and Julien P. H. Verheyden Syntex Research, Palo Alto, CA 94304

Abstract. The synthesis of 4'-(hydroxymethyl)guanosine (7) and the phosphonate analogue 8 of guanylic_acid proceed from a common intermediate, $2', 3'-\underline{0}$ -isopropylidene- \underline{N}^2 -(monomethoxytrityl)-guanosine-5'-aldehyde (13).

INTRODUCTION

A number of guanosine nucleoside analogues³ such as acyclovir (1)⁴ and ganciclovir (2)⁵ have been reported to be active against herpes simplex virus. For activity, these compounds are converted to their corresponding monophosphates by a viral thymidine kinase. Host kinases phosphorylate the monophosphates to triphosphates which in turn inhibit the viral DNA polymerase and thus virus replication. In as much as these analogues are not substrates for host kinases in the initial phosphorylation, selectivity is realized at this step.

Additionally, nucleotide analogues which may bypass the activation step by the viral thymidine kinase can exert an antiviral effect against viruses which lack this enzyme. An example of such a compound is ganciclovir phosphonate derivative 3 which is active in

vitro 7 and in vivo 8 against cytomegalovirus, a virus that does not encode a kinase. Even though phosphonate 3 bypasses the potential selectivity incurred by the kinase step, the compound showed very low toxicity in tissue culture indicating that such a nucleotide can be highly selective, presumably at the DNA polymerase level. Since the report of 3, a PFA derivative of guanosine 9 and HPMPA 9 0, an especially broad-spectrum antiviral agent, have been described as antiviral nucleotide analogues.

The activity of acyclic guanosine nucleoside and nucleotide analogues and the fact that the riboside derivative ribavirin (6) at least in part exerts its broad-spectrum activity as a guanosine analogue 11 prompted us to prepare guanosine derivatives 4'-(hydroxymethyl)guanosine (7) and the phosphonate analogue 8 of guanylic acid (9).

$$H_2N$$
 H_2N
 H_3N
 H_3N

RESULTS AND DISCUSSION

The chemistry utilized for the synthesis of 7 and 8 is based on previous work at this Institute directed towards the preparations of 4'-substituted nucleosides 12 and nucleoside phosphonates. 13 Both classes of compounds were approached via nucleoside 5'-aldehyde intermediates. For this guanosine series, we utilized the monomethoxytrityl protecting group at the $\underline{\text{N}}^2$ position, which we have previously found protects and solubilizes guanosine derivatives. 7,14

Thus, $5'-\underline{0}$ -acetyl-2', $3'-\underline{0}$ -isopropylideneguanosine ($\mathbf{10}$)¹⁵ (Scheme 1) was monomethoxytritylated to give $\mathbf{11}$ which in turn was deacetylated (NH₄OH/MeOH) furnishing alcohol $\mathbf{12}$ (quantitative). Alcohol $\mathbf{12}$ was suitably protected so that Moffatt oxidation¹⁶ afforded a quantitative yield of key intermediate aldehyde $\mathbf{13}$.

Reaction of the 5'-aldehyde 13 with paraformaldehyde in aqueous NaOH gave hydroxymethyl derivatives 14 and 15 in the low yields of 10% and 9%, respectively. The formation of methylene derivative 15 was

Scheme 1

not unexpected having been observed with a mechanism proposed previously for a similar reaction on \underline{N}^6 -benzoyl-2',3'- $\underline{0}$ -isopropylideneadenosine 5'-aldehyde. Complete deprotection of 14 with aqueous trifluoroacetic acid afforded target compound 7 in 94% yield. Alternatively, the methylene derivative 16 was prepared by the treatment of 15 with aqueous acetic acid.

Aldehyde 13 was also utilized for the synthesis of guanylic acid phosphonate analogue 8 (Scheme 2). Condensation of 13 with diphenyl tributylphosphoranylidene methylphosphonate 17 in tetrahydrofuran gave unsaturated phosphonate 17 which was directly reduced with diimide furnishing 18 in 25% yield. Transesterification of 18 with sodium benzyl oxide gave dibenzyl ester 19 (61%). Acetic acid hydrolysis of 19 to 20 followed by transfer hydrogenation furnished nucleotide analogue 8 in 29% yield after recrystallization from water.

Scheme 2

Analogues 7 and 8 were found to be inactive when tested in vitro against herpes simplex virus types 1 and 2, parainfluenza 3, and respiratory syncytial virus.

EXPERIMENTAL

Nuclear magnetic resonance spectra were recorded on a Bruker WM-300 (¹H NMR, 300 MHz) and chemical shifts are reported in parts per million downfield from internal tetramethylsilane. Ultraviolet spectra were recorded on a Hewlett Packard 8450A spectrometer, and spectroscopic data and elemental analyses were obtained by Syntex Analytical Research. All chromatographic purifications were carried out on silica gel. Melting points were determined on a hot-stage microscope and are corrected.

2',3'-0-Isopropylidene-N²-(monomethoxytrityl)guanine (12). A solution of 10 (11.60 g, 31.8 mmol), monomethoxytritylchloride (13.3g, 48 mmol), triethylamine (8.9 mL, 64mmol), and 4-dimethylaminopyridine (0.19g, 1.5 mmol) in DMF (150 mL) was heated at 50 $^{\circ}$ C for 1.5 h, and then methanol (10 mL) was added. The solution was evaporated to dryness. The residue was dissolved in ethyl acetate, washed with saturated aqueous $NaHCO_3$, dried over Na_2SO_4 , and evaporated to dryness. The residue was taken up in a mixture of methanol (100 mL) and conc. NH_AOH (40 mL), and the solution was stirred at room temperature for 20 h. Evaporation of the reagents left a syrup which was chromatographed (1:14 methanol/dichloromethane) to give 20 g (100%) of 12 as a foam; UV λ_{max} (methanol) 277 nm (ϵ 13500), 260 (15100); 1 H NMR (Me₂SO-d₆) δ 10.67 (s, broad, 1H, NH), 7.82 (s, 1H, H-8), 7.64 (s, broad, 1H, NH), 7.16-7.35 (m, 12H, aromatic), 6.86 (d, J = 9Hz, 2H aromatic), 5.36 (d, J = 4Hz, 1H, H-1'), 4.98 (t, J = 5 Hz, 1H, OH), 4.72 (dd, J = 4 and 6 Hz, 1H, H-2'), 4.37 (dd, J = 2.5 and 6Hz, 1H, H-3'), 3.96 (m, 1H, H-4'), 3.71 (s, 3H, OCH_3), 3.30 (m, 2H, H-5'), 1.35 (s, 3H, CH_3), 1.20 (s, 3H, CH_3). Anal. Calcd for $C_{33}H_{33}N_{5}O_{6} \cdot 1.5H_{2}O$ (622.68): C, 63.65; H, 5.83; N, 11.25. Found: C, 63.56; H, 5.86; N, 11.28.

4'-Hydroxymethyl-2',3'-Q-isopropylidene- N^2 -(monomethoxytrityl) quanosine (14) and 4'-hydroxymethyl-2',3'-0-methylene-N²-(monomethoxytrityl)quanosine (15). A solution of 12 (6.23 g, 10 mmol), dicyclohexylcarbodiimide (8.25 g, 40 mmol) and methylphosphonic acid (0.86 g, 10 mmol) in DMSO (100 mL) was stirred at $18 \, ^{\circ}\text{C}$ for 4 h and then at room temperature for 18 h. The resulting suspension was cooled to 18 °C, and oxalic acid dihydrate (125 mg) as a solution in methanol (6 mL) was added. The suspension was filtered, and the filtrate was evaporated to dryness in a Kugelrohr apparatus (60 °C/1 mm) to give aldehyde 13. A mixture of the residue in 37% aqueous paraformaldehyde (40 mL), 1 N sodium hydroxide (40 mL), THF (250 mL) and water (250 mL) was kept at room temperature for 48 h. Ammonium chloride (4 g) was then added and the solution evaporated to approximately 200 mL. The resulting mixture was extracted with dichloromethane. The organic phase was dried over Na_2SO_4 and evaporated to dryness. The residue was chromatographed (1:14 methanol/dichloromethane) to give 0.621 g (10%) of 14 and 0.556 g (9%) of the more polar 15. Both products were isolated as amorphous solids.

14: UV λ_{max} (methanol) 277 nm (ϵ 14200), 261 (15800); 1 H NMR (Me₂SO-d₆) δ 10.61 (s, broad, 1H, NH), 7.87 (s, 1H, H-8), 7.62 (s, broad, 1H, NH), 7.16-7.35 (m, 12H, aromatic), 6.84 (d, J = 9Hz, 2H, aromatic), 5.31 (d, J = 4.7 Hz, 1H, H-1), 4.96 (t, J = 5 Hz, 1H, OH), 4.80 (dd, J = 5 Hz, 1H, H-2'), 4.59 (t, J = 5 Hz, 1H, OH), 4.53 (d, J = 5 Hz, 1H, H-3'), 3.71 (s, 3H, OCH₃), 3.15-3.62 (m, 4H, CH₂OH), 1.26 (s, 3H, CH₃), 1.20 (s, 3H, CH₃)). Anal. Calcd for $C_{34}H_{35}N_{5}O_{7} \cdot H_{2}O$ (643.71): C, 63.44; H, 5.79; N, 10.88. Found: C, 63.27; H, 5.77; N, 10.80.

15: UV λ_{max} (methanol) 277 nm (ϵ 14200), 260 (16000); ${}^{1}\text{H}$ NMR (Me₂SO-d₆) δ 10.66 (s, broad, 1H, NH), 7.82 (s, 1H, H-8), 7.63 (s, broad, 1H, NH), 7.17-7.35 (m, 12H, aromatic), 6.86 (d, J=9Hz, 2H, aromatic), 5.37 (d, J = 4.2 Hz, 1H, H-1'), 4.90 (t, J = 5 Hz, 1H, OH), 4.88 (s, 2H, OCH₂O), 4.79 (dd, J = 4 and 6 Hz, 1H, H-2'), 4.62 (t, J = 5 Hz, 1H, OH), 4.25 (d, J = 6Hz, 1H, H-3'), 3.72 (s, 3H, OCH₃), 3.10-3.60 (m, 4H, CH₂OH). Anal. Calcd for $C_{32}H_{31}N_{5}O_{7}\cdot H_{2}O$ (615.65): C, 62.43; H, 5.40; N, 11.38. Found: C, 62.44; H, 5.40; N, 11.37.

4'-Hydroxymethylguanosine (7). A solution of **14** (0.621 g, 0.99 mmol) in 50% aq trifluoroacetic acid (7 mL) was kept at room temperature for 1 h and then evaporated to dryness. The residue was triturated with ethyl acetate and hexane to give 0.293 g (94%) of **7** as a beige powder. An analytical sample was obtained by recrystallization from water: mp >300 °C; UV λ_{max} (0.1 N HCl) sh 279 nm (\$\pi\$ 8500), 257 (12200), (0.1 N NaOH) 267 (11200); ¹H NMR (Me₂SO-d₆/D₂O) δ 7.96 (s, 1H, H-8), 5.73 (d, J = 7Hz, 1H, H-1'), 4.61 (dd, J = 5 and 7 Hz, 1H, H-2'), 4.15 (d, J = 5 Hz, 1H, H-3'), 3.50-3.80 (m, 4H, CH₂OH). Anal. Calcd for C₁₁H₁₅N₅O₆ · 0.5 H₂O (322.28): C, 41.00; H, 5.00; N, 21.73. Found: C, 41.00; H, 5.01, N, 21.80.

4'-Hydroxymethy1-2',3'-Q-methyleneguanosine (16). A solution of 15 (556 mg, 0.93 mmol) in 80% aqueous acetic acid was heated at 80 °C for 2 h and then evaporated to dryness. The residue was triturated with ethyl acetate and then recrystallized from methanol to give 78 mg (26%) of 16 mp >300 °C; UV λ_{max} (methanol) sh 270mm (ε 10100), 254 (14200); ¹H NMR (Me₂SO-d₆) δ 10.40 (s, broad, 1H, NH), 8.00 (s, 1H, H-8), 6.52 (s, broad, 2H, NH₂), 5.90 (d, J = 4 Hz, 1H, H-1'), 5.27 (dd, J = 4 and 6 Hz, 1H, H-2'), 5.22 (t, J = 5 Hz, 1H, OH), 5.19 and 5.06 (s, 1H, OCH₂O) 4.77 (t, J = 5 Hz, 1H, OH), 4.72 (d, J = 6 Hz, 1H, H-3'), 3.45-3.72 (m, 4H, CH₂OH). Anal. Calcd for $C_{12}H_{15}N_5O_6 \cdot 0.33 H_2O$ (331.29); C, 43.51; H, 4.77; N, 21.14. Found: C, 43.64; H, 4.77; N, 21.19.

9-[5,6-dideoxy-6-Diphenoxyphosphinyl-2,3-Q-isopropylidene- β -D-ribo-hex-5-enofuranosyl]- \underline{N}^2 -monomethoxytritylguanine (17). A solution of 13 (10 mmol, prepared as described above), diphenyl tributylphosphoranylidenemethylphosphonate (7.28 g, 15 mmol), and triethylamine (2.8 mL, 20 mmol) in THF (200 mL) was kept at room temperature for 72 h. After evaporation to dryness, the residue was dissolved in dichloromethane, washed with water, dried over Na_2SO_4 and evaporated to dryness. The residue was chromatographed three times (1:24 methanol/dichloromethane) to give 2.75 g of 17 still contaminated with a small amount of tributylphosphine oxide. 1 H NMR (Me_2SO-d_6) δ 10.76 (s, broad, 1H, NH), 7.77 (s, 1H, H-8), 7.69 (s, broad, 1H, NH), 7.10-7.45 (m, 22H, aromatic), 6.87 (d, J = 9 Hz, 2H,

aromatic), 6.80 (d, J = 2 Hz, 1H, H-1'), 6.71 (ddd, $J_{5'6'}$ = 17.4, $J_{5'P}$ = 24.2, $J_{5'4'}$ = 4.7Hz, 1H, H-5'), 4.98 (dd, J = 2 and 6 Hz, 1H, H-2'), 5.61 (ddd, $J_{6'5'}$ = 17.4, $J_{6',P}$ = 22.5, $J_{6',4'}$ = 1.7 Hz, 1H, H-6'), 4.67 (m, 1H, H-4'), 3.99 (dd, J = 3 and 6 Hz, 1H, H-3'), 3.70 (s, 3H, OCH₃), 1.34 (s, 3H, CH₃), 1.23 (s, 3H, CH₃).

9-[5,6-Dideoxy-6-diphenoxyphosphiny1-2,3-Q-isopropylidene-β-Dribo-hexofuranosyl]-N²-monomethoxytritylguanine (18). A solution of impure 17 from above (2.10 g, 2.5 mmol), potassium azidodicarboxylate (5.5 g, 2.8 mmol), and acetic acid (3.2 mL) in pyridine (70 mL) was kept at room temperature for 4 days and then evaporated to dryness. The residue was dissolved in dichloromethane, washed with water, dried over Na₂SO₄, and then evaporated to dryness. The residue was chromatographed (1:24 methanol/dichloromethane) and the product recrystallized from ethanol to give 0.514g (25%) of 18: mp 163-165 °C; UV λ_{max} (methanol) 277 nm (ϵ 14000), 261 (16600); ¹H NMR (Me_2SO-d_6) & 10.70 (s, broad, 1H, NH), 7.80 (s, 1H, H-8), 7.63 (s, broad, 1H, NH), 7.10-7.45 (m, 22H, aromatic), 6.85 (d, J = 9 Hz, 2H, aromatic), 5.51 (d, J = 3 Hz, 1H, H-1'), 4.94 (dd, J = 3 and 6 Hz, 1H, H-2'), 4.00 (m, 2H, H-3', H-4'), 3.70 (s, 3H, OCH₃), 1.60-2.15 (m, 4H, H-5', H-6'), 1.38 (s, 3H, CH_3), 1.20 (s, 3H, CH_3). Anal. Calcd for $C_{46}H_{44}N_5O_8P$ (825.86): C, 66.90; H, 5.37; N, 8.48; P, 3.75. Found: C, 66.93; H, 5.51; N, 8.54; P, 3.93.

9-[6-Dibenzyloxyphosphinyl-5,6-dideoxy-2,3-Q-isopropylidene-b-D-ribo-hexofuranosyl]-N²-monomethoxytritylguanine (19). To a suspension of NaH (0.25 g, 50%, 5.2 mmol; prewashed with hexane) and benzyl alcohol (1.1 mL, 10.7 mmol) in dry DMSO (15 mL) was added a solution of 18 (1.04 g, 1.13 mmol) in DMSO (10 mL). After 3 min, the resulting solution was diluted with ethyl acetate, washed twice with 1% aqueous NH₄Cl, dried over Na₂SO₄, and evaporated to dryness. The residue was chromatographed (1:19 methanol/dichloromethane) to give 0.656 g (61%) of 19 as an amorphous solid: UV (methanol) λ_{max} 277 nm (ϵ 14000), 263 (15600); ¹H NMR (Me₂SO-d₆) δ 10.68 (s, broad, 1H, NH), 7.75 (s, 1H, H-8), 7.61 (s, broad, 1H, NH), 7.12-7.41 (m, 22H, aromatic), 6.84 (d, J = 9 Hz, 2H, aromatic), 5.44 (d, J = 3Hz, 1H, H-1'), 4.85-5.03 (m, 5H, H-2', benzylic), 3.87 (m, 2H, H-3', H-4'),

3.69 (s, 3H, OCH₃), 1.40-1.80 (m, 4H, H-5', H-6'), 1.35 (s, 3H, CH₃), 1.19 (s, 3H, CH₃). Anal. Calcd for $C_{48}H_{48}N_5O_8P$ (853.92): C, 67.52; H, 5.67, N, 8.20. Found: C, 67.33; H, 5.67; N, 8.03.

9-[6-Dibenzyloxyphosphinyl]-5,6-dideoxy-2,3-Q-isopropylidene- β -D-<u>ribo</u>-hexofuranosyl]guanine (20). A solution of 19 (446 mg, 0.52 mmol) in 80% acetic acid was heated at 80 °C for 10 h and then evaporated to dryness. The residue was chromatographed (1:5 methanol/dichloromethane) to give 160 mg (57%) of 20 as a white solid: 1 H NMR (Me₂SO-d₆) δ 10.72 (s, broad, 1H, NH), 7.84 (s, 1H, H-8), 7.35 (s, 10H, phenyl), 6.54 (s, broad, 2H, NH₂), 5.65 (d, J = 2.7 Hz, 1H, H-1'), 5.44 (d, J = 5.8 Hz, 1H, OH), 5.15 (d, J = 5.2 Hz, 1H, OH), 5.00 (m, 4H, benzylic), 4.44 (m, 1H, H-2'), 3.8-3.9 (m, 2H, H-3', H-4'), 1.90 (m, 4H, H-5', H-6').

9-[5,6-Dideoxy-6-dihydroxyphosphiny]-β-D-ribo-hexofuranosy] guanine (8). A suspension of 20 (61.0 mg, 0.11 mmol) and 20% $Pd(OH)_2/C$ (30 mg) in cyclohexene (1 mL), ethanol (2 mL) and water (2 mL) was heated at reflux for 4h and then filtered through Celite. The filtrate was evaporated to dryness. The residue was crystallized from water to give 20 mg (50%) of 8: mp browns gradually at 200 °C, UV λ_{max} (0.1N HCl) sh 280 nm (ε 7890), 256 (11600), (0.1N NaOH) 267 (7890); 1 H NMR (D₂O) δ 8.33 (s, 1H, H-8), 5.91 (d, J = 5 Hz, 1H, H-1'), 5.80 (m, buried under HOD, 1H, H-2'), 4.28 (dd, J = 5 Hz, 1H, H-3'), 4.17 (m, 1H, H-4'), 1.60-2.10 (m, 4H, H-5', H-6'). Anal. Calcd for $C_{11}H_{16}N_5O_7P\cdot1.5H_2O$ (388.28): C, 34.03; H, 4.93; N, 18.04. Found: C, 33.85; H, 4.75; N, 17.97.

Acknowledgement. We thank Drs. John G. Moffatt and Gordon H. Jones for their advice. The assistance of Dr. M. Maddox, Ms. J. Nelson and Ms. L. Kurz in obtaining and interpreting NMR spectra is appreciated.

REFERENCES

- Contribution 216 from the Institute of Bio-Organic Chemistry.
- Current address: Bristol-Myers Pharmaceutical Research and Development Division, Wallingford, CT 06492-7660.

- Remy, R. J.; Secrist, J. A. <u>Nucleosides and Nucleotides</u>, 1985, 4, 411. Chu, C. K.; Cutler, S. J. <u>J. Heterocyclic Chem.</u> 1986, 23, 289.
- Schaeffer, H. J.; Beauchamp, L.; Miranda, P.; Elion, G.; Bauer, D. J.; Collins, P. Nature (London) 1978, 272, 583.
- (a) Martin, J. C.; Dvorak, C. A.; Smee, D. F.; Matthews, T. R.; Verheyden, J. P. H. J. Med. Chem. 1983, 26, 759. (b) Schaeffer, H. J. in Nucleosides, Nucleotides and Their Biological Applications; Rideout, J. L.; Henry, D. W.; Beecham, L. M.; Eds.; Academic Press: New York, 1983; pp 1-17. (c) Ashton, N. J.; Karkas, J. D.; Field, A. K.; Tolman, R. L. Biochem. Biophys. Res. Comm. 1982, 108, 1716, (d) Smith, K. O.; Galloway, K. S.; Kennell, W. L.; Ogilvie, K. K.; Radatus, B. K. Antimicrob. Agents Chemother. 1982, 22, 55.
- 6. Robins, R. K. Pharm. Res. 1984, 11.
- 7. Prisbe, E. J.; Martin, J. C.; McGee, D. P. C.; Barker, M. F.; Smee, D. F.; Duke, A. E.; Matthews, T. R.; Verheyden, J. P. H. J. Med. Chem. 1986, 29, 671.
- Duke, A. E.; Smee, D. F.; Chernow, M.; Boehme, R.; Matthews, T. R. Antiviral Research 1986, 6, 299.
- Vaghefi, M. M.; McKernan, P. A.; Robins, R. K. <u>J. Med. Chem.</u> 1986, 29, 1389.
- 10. De Clercq, E.; Holy, A.; Rosenberg, I.; Sakuma, T.; Balzarini, J.; Maudgal, P. C.; Nature **1986**, 323, 464.
- Gilbert, B. E.; Knight, V. <u>Antimicrob. Agents Chemother.</u> 1986, 30, 201.
- (a) Youssefyeh, R. D.; Verheyden, J. P. H.; Moffatt, J. G. J. Org. Chem. 1979, 44, 1301. (b) Jones, G. H.; Taniguchi, M.; Tegg, D.; Moffatt, J. G. J. Org. Chem. 1979, 44, 1309.
- 13. Jones, G. H.; Moffatt, J. G. <u>J. Amer. Chem. Soc.</u> **1968**, <u>90</u>, 5337.
- 14. (a) Nerenberg, C.; McClung, S.; Martin, J.; Fass, M.; La Fargue, J.; Kushinsky, S. <u>Pharmaceutical Research</u> 1986, 3, 112, (b) Martin, J. C.; McGee, D. P. C.; Jeffrey, G. A.; Hobbs, D. W.; Smee, D. F.; Matthews. T. R.; Verheyden, J. P. H. <u>J. Med. Chem.</u> 1986, 29, 1384.
- Dimitrijevitch, S. D.; Verheyden, J. P. H.; Moffatt, J. G. J. Org. Chem. 1979, 44, 400.
- Pfitzner, K. E.; Moffatt, J. G. <u>J. Amer. Chem. Soc.</u> 1965, <u>87</u>, 5661, 5670.
- 17. Jones, G. H.; Hanamura; E. K.; Moffatt, J. G. <u>Tetrahedron Lett.</u> 1968, 5173.